Views and Perspectives

The Migraine Attack as a Homeostatic, Neuroprotective Response to Brain Oxidative Stress: Preliminary Evidence for a Theory

Jonathan M. Borkum, PhD

Background.—Previous research has suggested that migraineurs show higher levels of oxidative stress (lipid peroxides) between migraine attacks and that migraine triggers may further increase brain oxidative stress. Oxidative stress is transduced into a neural signal by the TRPA1 ion channel on meningeal pain receptors, eliciting neurogenic inflammation, a key event in migraine. Thus, migraines may be a response to brain oxidative stress.

Results.—In this article, a number of migraine components are considered: cortical spreading depression, platelet activation, plasma protein extravasation, endothelial nitric oxide synthesis, and the release of serotonin, substance P, calcitonin gene-related peptide, and brain-derived neurotrophic factor. Evidence is presented from in vitro research and animal and human studies of ischemia suggesting that each component has neuroprotective functions, decreasing oxidant production, upregulating antioxidant enzymes, stimulating neurogenesis, preventing apoptosis, facilitating mitochondrial biogenesis, and/or releasing growth factors in the brain. Feedback loops between these components are described. Limitations and challenges to the model are discussed.

Conclusions.—The theory is presented that migraines are an integrated defensive, neuroprotective response to brain oxidative stress.

Key words: migraine, oxidative stress, growth factor, neuroprotective, homeostasis, regenerative

Abbreviations: AMPK adenosine monophosphate-activated protein kinase, BACE1 β-site amyloid precursor protein cleaving enzyme 1, BDNF brain-derived neurotrophic factor, bFGF basic fibroblast growth factor, CGRP calcitonin gene-related peptide, CREB cyclic AMP-response element-binding transcription factor, CSD cortical spreading depression, DRN dorsal raphe nucleus, eNOS endothelial nitric oxide synthase, ERK extracellular signal regulated kinase, FGF fibroblast growth factor, GDNF glial cell line-derived neurotrophic factor, IGF-1 insulin-like growth factor-1, iNOS inducible nitric oxide synthase, MAPK mitogen activated protein kinase, NADPH nicotinamide adenine dinucleotide phosphate, NF-κB nuclear factor-kappaB, NGF nerve growth factor, Nrf2 nuclear factor (erythroid-derived 2)-like 2, PAF platelet activating factor, PDGF platelet-derived growth factor, PI3K phosphoinositide 3-kinase, PKC protein kinase C, PLC-γ1 phospholipase C-gamma1, PPE plasma protein extravasation, pTrkB phosphorylated tropomyosin-related kinase B, SSRI selective serotonin reuptake inhibitor, TGF-β1 transforming growth factor-beta1, tPA tissue plasminogen activator, TrkB tropomyosin-related kinase B, TRPA1 transient receptor potential ankyrin-1, VEGF vascular endothelial growth factor

(Headache 2018;58:118-135)

From the Department of Psychology, University of Maine, Orono, ME, USA (J.M. Borkum); Health Psych Maine, Waterville, ME, USA (J.M. Borkum) email: jborkum@hpmaine.com

Address all correspondence to J. M. Borkum, Department of Psychology, University of Maine, 301 Little Hall, Orono, ME 04469-5782, USA, email: jborkum@hpmaine.com

Accepted for publication August 13, 2017.

Migraines are a multifaceted neurological disorder comprised of head pain, nausea or vomiting, and/or painful avoidance of light and sound, and aggravated by or causing avoidance of routine physical activity. Five lifetime attacks are required for diagnosis (two for migraine with aura) because isolated migraine attacks can affect nearly anyone, and resemble secondary headaches due to such threats to the brain as meningitis and carbon monoxide poisoning.

Current understandings of migraine focus on cortical hyperexcitability, 2,3 activation of the trigeminovascular system.⁴ and dysregulation of brainstem regions involved in antinociception and vascular control.⁵⁻⁷ However, there is also an additional, minor paradigm: that migraine attacks are an adaptive, compensatory reaction. Thus, Loder observed that their partial heritability and high prevalence make it likely that migraines confer an evolutionary advantage, such as a defense against threats to the nervous system.8 Goadsby suggested that the vasodilation of migraines is a "neural protection system" against excessive vasoconstriction.9 And Cortelli and Montagna noted that the visceral pain of migraine and the immobility, sleep, and aversion to sensory stimulation that it incites seem well positioned to ameliorate a brain energy imbalance. 10

In this article, we will propose and consider the merits of a particular adaptive theory of migraine attacks: that they are an integrated defensive response to oxidative threats to the brain. The goal is to provide a starting point for discussion.

MIGRAINES AND OXIDATIVE STRESS

Vulnerability to migraines is not completely understood but may be in part related to higher levels of lipid peroxides found in plasma, platelets, and urine of migraineurs between attacks. Lipid peroxides are oxidation products and thus a biomarker of oxidative stress. The reason for higher levels interictally in migraineurs is unclear but may reflect lower activity of certain antioxidant enzymes, and/or increased activity of oxidant-generating

Conflicts of Interest: None.

Sources of Financial Support: None.

vasoconstrictors such as angiotensin,^{16,17} endothelin-1,¹⁸ or urotensin-2.¹⁹ Alternatively, the source may be cortical hyperexcitablity,² perhaps resulting in excessive oxidant production as a byproduct of the high metabolic rate.²⁰ Moreover, migraines may be associated with mitochondrial defects,²¹⁻²⁵ which tend to increase production of oxidants²⁶ and downregulate antioxidant enzymes.²⁷ Of course, more than one of these processes may be involved; oxidative stress may be a final common pathway for a number of unfavorable conditions in the brain.

Lipid peroxides are plausible signaling molecules for oxidative stress because they have long half-lives, and no net charge, and thus are able to diffuse, reaching receptors. In fact, there appear to be dedicated receptors for oxidative stress on nociceptive nerve endings in the perivascular meninges. Specifically, TRPA1 ion channels (transient receptor potential ankyrin type 1) open when certain of its cysteine residues are oxidized, admitting calcium ions and transducing the oxidizing conditions into a neural signal. Moreover, in animal models, activation of the TRPA1 ion channel is sufficient to trigger neurogenic inflammation, a key component of migraines. In the provided stress of the provided stress of the plausible stress of the plau

Migraine triggers may be exposures that further increase oxidative stress.^{20,31} For certain putative triggers, hydrogen peroxide is a byproduct of detoxification, for example when tyramine is metabolized by monoamine oxidase or when alcohol is processed by cytochrome P450-2E1. Other triggers appear to cause oxidant production by the microglia, neurons, and astrocytes as part of host defense. These triggers include infection, air pollution (particulate matter), aspartame, hypoxia, hypoglycemia and, potently, psychosocial stress,³² which in the natural world may connote a high probability of imminent attack and infection. Other triggers may raise oxidant production by increasing the rate of oxidative phosphorylation, potentially extending to excitotoxicity. These may include, in susceptible individuals, monosodium glutamate, aspartame (via its aspartic acid moiety), hypoglycemia, and perhaps exposure to noise and mental overwork. For still other triggers, the oxidant production may arise from mitochondrial dysfunction. Alcohol, formate (a metabolite of aspartame),

and β -phenylethylamine (found in chocolate) fall in this category.²⁰

Thus, migraineurs may have a higher level of oxidative stress between attacks and migraine triggers may increase oxidant production further. The TRPA1 ion channel may provide a mechanistic link to migraines, eliciting neurogenic inflammation in response to oxidative stress.

Being able to detect and respond to oxidative stress would be adaptive, as the brain's polyunsaturated (easily oxidized) neuronal membranes, the presence of transition metal ions that can catalyze free radical reactions, high metabolic rate, and relatively weak antioxidant defenses make it uniquely exposed and susceptible to damage from oxidants.³³ In turn, this damage may be an early step in neurodegenerative diseases such as Alzheimer's or Parkinson's.³⁴

However, if migraines are plausibly a response to brain oxidative stress, what type of response are they? In this article, we will explore the possibility that they are a coordinated set of mechanisms that deliver antioxidants and growth factors to the brain, reverse oxidative stress, prevent apoptosis, and support the survival, proliferation, development, and complex architecture of neurons. Indeed, there is evidence that numerous components of migraine physiology may be designed to protect and repair the brain.

Components of Migraine.—Before considering this theory, let us first review the main components of a migraine attack, drawing on current understanding of migraine pathophysiology. The concern here is not for the precise order or timing of the components, but simply a delineation of the components themselves.

Episodic migraine is classically thought to be a functional neurovascular disorder of the brain.³⁵ Initially, in migraine aura a wave of depolarization and hyperemia, followed by oligemia and suppression of spontaneous neural activity, propagates geographically over the cortex, producing such aura symptoms as scintillations and scotomas (cortical spreading depression; CSD). In migraine without aura, the situation is less clear but may involve "silent" CSD in nonsensory, nonmotor regions of the cortex. The high concentration of glutamate and potassium ions from CSD is thought to irritate

the small diameter nociceptive fibers surrounding the cerebral blood vessels, venous sinuses, and dura mater. (As we have seen, it is likely that nociception can be initiated also by the products of reactive oxygen and nitrogen species.) This causes the fibers to release substance P and calcitonin generelated peptide (CGRP),³⁶ initiating neurogenic inflammation of the dura mater, with mast cell degranulation, vasodilation, hyperemia, plasma protein extravasation (PPE) from the postcapillary venules, platelet aggregation in the venules, and possibly further sensitization of the nociceptors.³⁷ Activation is conveyed by these nociceptive fibers to the trigeminal nucleus caudalis and the thalamus, where sensitization of second and third order neurons, perhaps in part through CGRP,³⁸ contributes to the pain of migraine.

The trigeminal nucleus also conveys the nociceptive signal to a range of nuclei in the brainstem, hypothalamus, and basal ganglia,³⁹ likely including the dorsal raphe nucleus (DRN). Platelets and presumably the DRN release serotonin that, via 5-HT_{2B} receptors in the blood vessels, activate endothelial nitric oxide synthase (eNOS), increasing the production of nitric oxide. In turn, eNOS⁴⁰ and platelet activation⁴¹ likely cause release of brainderived neurotrophic factor (BDNF) from the endothelium and platelets, respectively.

Aspects of Neuroprotection.—There are signaling pathways within neurons that protect them against apoptosis in oxidizing conditions, suppress the production of oxidants and facilitate their detoxification, support the development of synapses and the elaborate arborization of dendrites, repair neurons after injury, and facilitate neurogenesis—the birth of new neurons.

The most important of these pathways are: 42-44

- A pathway from phosphoinositide 3-kinase (PI3K) to Akt (protein kinase B), which suppresses certain transcription factors (p53, forkhead FKHRL1) and facilitates others (nuclear factor-kappaB/NF-κB)
- A pathway through extracellular signal regulated kinases (ERK), a type of mitogen-activated protein kinase (MAPK), which ultimately activates

the cyclic AMP-response element binding transcription factor (CREB), and

 A pathway through phospholipase C-γ1 (PLCγ1), which activates a number of enzymes including protein kinase C (PKC).

These signaling cascades are set in motion by various neurotrophic factors such as BDNF, nerve growth factor (NGF), glial cell line-derived neurotrophic factor (GDNF), and basic fibroblast growth factor (bFGF or FGF2).⁴³ In particular, BDNF can activate all three signaling pathways.^{43,44}

Thus, given that migraines may be a response to increased levels of oxidative stress in the brain, in what follows we will explore whether any of the components of migraine can deliver antioxidants to the brain, decrease the production of oxidants, and/ or activate the signaling pathways of neural protection and repair. In doing so, we will draw on in vitro data and on animal and, where available, clinical, studies of ischemia. The appropriateness of using ischemia as a model condition will be discussed at the end of this article.

COMPONENTS OF MIGRAINE THROUGH THE LENS OF NEUROPROTECTION

Platelet Activation.—Migraines involve platelet activation and aggregation. In particular, levels of the highly potent platelet activating factor (PAF) rise in the internal jugular venous blood during spontaneous migraine attacks. In turn, platelets release serotonin, which may sensitize nociceptive fibers around pial and intracerebral blood vessels; proinflammatory cytokines, contributing to neurogenic inflammation; and nitric oxide, which is vasodilatory. In particular, levels of the highly potential properties around pial and intracerebral blood vessels; proinflammation; and nitric oxide, which is vasodilatory.

Classically, of course, platelets are responsible for hemostasis. However, once activated, platelets also deliver a range of growth factors that promote tissue healing and regeneration. These include vascular endothelial growth factor (VEGF), FGF, platelet-derived growth factor (PDGF), transforming growth factor- β 1 (TGF- β 1), NGF, and BDNF. These molecules, in turn, switch on (phosphorylate) growth-oriented cell signaling cascades such as the ERK and Akt pathways. By these

means, platelets create a healing environment at a site of injury. 50

This appears to be true in the brain as well. Acting in part through VEGF, FGF, and PDGF by shedding microparticles, activated platelets induce neural stem cell survival, proliferation, and differentiation into neurons and glia, as well as stimulating angiogenesis, in vitro, 50 and are neuroprotective and neuroregenerative in an in vivo model of cerebral ischemia. 51 This has led to the theory that platelets participate in the housekeeping of nervous tissue. 52

Plasma Protein Extravasation.—In extravasation, proteins from blood serum leak into cerebrospinal fluid. The function of PPE in migraine is unclear. However, substance P, which elicits PPE, protects neurons from apoptosis in vitro.⁵³ Further, the most abundant serum protein is albumin, which is an antioxidant. In serum, 70% of the capacity to neutralize free radicals is from albumin specifically.⁵⁴ Logically, after extravasation into cerebrospinal fluid during migraine, albumin may play an antioxidant role in the brain.

In animal models of ischemic stroke, intravenous administration of human albumin leads to its extravasation and uptake by cerebral neurons. These neurons, in turn, seem to be protected from damage^{55,56} and cortical infarct volume is reduced.⁵⁷ Albumin, which carries polyunsaturated fatty acids, seems to help in the repair of neuronal cell membranes.⁵⁸ Further, albumin administered in vitro to astrocytes is taken up by the megalin receptor and causes the production of oleic acid, which is neurotrophic, facilitating the differentiation of neurons.^{55,59,60}

Unfortunately, in a large clinical trial of ischemic stroke, high dose IV albumin did not improve outcome, apparently because adverse events balanced out the treatment effect. Preliminary clinical trial data, however, suggest that IV albumin may help prevent delayed cerebral ischemia and cerebral infarction following subarachnoid hemorrhage.

Production of Nitric Oxide.—An early event in migraine is activation of eNOS or NOS-3,⁶⁴ likely through the 5-HT_{2B} receptor.⁶⁵ The nitric oxide so produced contributes to the vasodilation in migraines and, by stimulating the release of substance P, to extravasation.⁶⁶

In addition, in the vasculature, nitric oxide downregulates the renin-angiotensin system by inhibiting the production of angiotensin-converting enzyme and angiotensin-II AT₁ receptors.⁶⁷ Angiotensin increases oxidant production by strongly activating the enzyme nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase)⁶⁷ and thus its downregulation by nitric oxide has an antioxidant effect.

Moreover, eNOS is thought to protect against neuronal damage under hypoxic or ischemic conditions. In part, this is by helping to maintain cerebral blood flow and in part, nitric oxide, acting through cyclic GMP and the Nrf2 (nuclear factor [erythroid-derived 2]-like 2) transcription factor, induces antioxidant enzymes and proteins including heme oxygenase-1 and sestrin2. Endothelial nitric oxide facilitates mitochondrial biogenesis and the production of ATP, to potentially correcting for mitochondrial dysfunction and impaired brain energetics as diatheses for migraine. And endothelial nitric oxide can also induce angiogenesis, and the proliferation and migration of neural progenitor cells, to components of tissue repair.

Endothelium-derived nitric oxide contributes to synaptic plasticity and long-term potentiation in the brain, ⁷⁶ in part because the blood vessel endothelium, in the presence of active (phosphorylated) eNOS, releases BDNF into the brain. ⁴⁰ Indeed, the blood vessel endothelium is a major source of cerebral BDNF, ⁷⁷ with endothelial cells producing BDNF at 50 times the rate of neurons. ⁷⁸ In turn, BDNF may enhance vasodilation and the release of nitric oxide in a positive feedback loop. ^{77,79} Thus, the paradigm that migraines are an integrated neurovascular phenomenon extends to neural protection and repair.

Brain-Derived Neurotrophic Factor.—BDNF is the most abundant and most studied of the trophic factors in the brain. So Serum levels of BDNF seem to rise during attacks. Moreover, platelet levels are lower in migraineurs, so suggesting that the platelets release BDNF as part of their activation during the migraine process. Expression of the gene encoding BDNF increases 15-fold 3 hours after CSD in the rat, while expression of the gene for its

receptor triples.⁸⁴ In vitro, BDNF is released from trigeminal ganglion neurons in response to stimulation by CGRP.⁸⁵

BDNF plays a key role in central sensitization⁸⁶ including, potentially, in migraine. However, it also appears to have a larger, neuroprotective role. Acting through the tropomyosin-related kinase B (TrkB) receptor, BDNF seems to preserve neural progenitor cells, prevent the degeneration of cortical neurons,^{87,88} promote the formation, maintenance, and plasticity of synapses,⁸⁸ and support the consolidation of memory from short-term to long-term stores.⁸⁷

At least in rats, BDNF is also involved in neural repair, promoting recovery from spinal cord injury and, in the brain, rescuing synapses and cognition after experimental transient ischemia ⁸⁹⁻⁹² and psychosocial stress. ⁹³ TrkB is found in blood vessels and its stimulation promotes angiogenesis, ⁸⁷ particularly relevant after ischemia. Neurogenesis from exercise, an enriched environment, or chronic treatment with antidepressants seems to depend on increased BDNF. ⁹⁴

Moreover, BDNF signaling sets in motion an array of antioxidant processes, including upregulation of the enzymes glutathione reductase and superoxide dismutases; sulfiredoxin, an antioxidant protein that protects mitochondria; and sestrin2, a protein that regenerates oxidized peroxyredoxins and upregulates the Nrf2 antioxidant transcription factor. BDNF also decreases the production of superoxide, a free radical, by limiting the leakage of electrons from the electron transport chain. 96

Thus, BDNF plays a large role in brain protection and repair. Not surprisingly, then, increasing BDNF levels has been proposed as a treatment strategy for Parkinson's disease, ⁹⁷ Huntington's chorea, ⁹⁸ and stroke rehabilitation. ⁹⁹

CGRP Release.—CGRP is a key effector molecule in migraine. ¹⁰⁰ Released by perivascular sensory fibers, it contributes to vasodilation, mast cell degranulation, neurogenic inflammation, and possibly peripheral pain sensitization in migraine. ¹⁰¹ As a neurotransmitter in the trigeminal nucleus caudalis, it contributes to central sensitization. ¹⁰²

In addition, however, CGRP reduces oxidative stress in the body, 103 in part by lowering the expression of the enzyme, NADPH oxidase, which produces superoxide, 104 and in part by increasing the activity of antioxidant enzymes. 103 Further, CGRP prevents apoptosis under oxidizing conditions. 105 Moreover, like activated platelets, CGRP facilitates wound healing and tissue repair in the body, in part by increasing the expression of VEGF, bFGF, and TGF- β . 101 Indeed, a loss of CGRP-containing sensory nerves may account for some of the impaired wound healing seen in diabetes. 101

In the endothelium, CGRP antagonizes the oxidant-inducing effects of angiotensin II much as does nitric oxide. ¹⁰¹ In the brain, CGRP expression in neurons is upregulated by axonal damage, infection, and inflammation ¹⁰⁶ and CGRP in turn activates astrocytes and microglia. ^{106,107} Moreover, CGRP is a strong vasodilator and, under ischemic conditions, activates the CREB transcription factor, and raises levels of bFGF and the protein Bcl-2, suppressing apoptosis of neurons. ^{108,109} Thus, CGRP is thought to be part of the central nervous system's response to ischemia, injury, and infection, ^{9,110-113} inducing repair. ¹⁰⁶

Accordingly, injection of CGRP directly into a cerebral ventricle improves learning and memory in rats, 114 increases neurogenesis and the expression of NGF in mice subjected to combined restraint and water immersion stressors, 115 and raises production of GDNF in a mouse model of amyotrophic lateral sclerosis. 106

Indeed, CGRP has been used in two controlled human trials to reverse persistent neurological deficits, attributed to vasospastic ischemia, after aneurysm surgery. In the larger trial, whose power was compromised by marked patient dropout due to systemic hypotension, the group receiving IV CGRP had numerically improved neurological outcome at 3 months (vs usual treatment) but this result was not statistically significant. More recent animal studies have focused instead on the downstream neurotrophic factor, bFGF, which has been shown to markedly reduce infarct volume, neurological deficit, and the impaired activity seen following cerebral ischemia-reperfusion in rats. In the second secon

Cortical Spreading Depression.—In CSD, a wave of intense activation and reactive hyperemia spreads geographically over the surface of the cortex, followed in its wake by a depression of spontaneous neural activity, thought to correspond to the scintillations and scotomas, respectively, of migraine aura. The propagation of CSD depends on the release of potassium ions and glutamate, which are thought to directly depolarize peripheral nociceptors, thereby initiating neurogenic inflammation and inducing a migraine. Further, CSD entails the production of oxidants, which can irritate nociceptors through the TRPA1 ion channel. Thus, CSD may be a key initiating event for the pain of migraine.

However, CSD is also a form of preconditioning, a minor stress that protects the brain against a subsequent, larger stressor. Thus, in animal models and tissue slices, CSD is known to limit the amount of damage caused by subsequent ischemia. In part, this seems to occur because CSD activates an enzyme, AMP-activated protein kinase (AMPK), that detects conditions of depleted energy and downregulates energy-demanding pathways. CSD also induces the transcription of a number of antioxidant and neuroprotective genes. Purther, an increase in uncoupling protein-5 reduces the production of a key oxidant, superoxide, by the mitochondria following CSD.

Moreover, the initial phase of CSD is a wave of depolarization and hyperemia. These increases in neural activity and blood flow should raise production of BDNF by neurons and the endothelium, respectively. In fact, BDNF mRNA and protein levels are markedly increased in the rat brain, ^{86,125-127} and long-term synaptic potentiation is strengthened, ¹²⁸ following CSD.

Release of Serotonin.—Electrophysiological, neurochemical, and platelet studies suggest that the interictal period in migraines is characterized by a serotonin deficit, which is rapidly reversed during a migraine attack. During a migraine, there is increased synthesis of serotonin, normalizing it to levels seen in nonmigraineurs, Iso increased firing in the region of the DRN^{6,131} potentially bathing the brain in serotonin, Sacrotonin is

Component	Direct Antioxidant	Induces Antioxidant Enzymes	↓ Oxidant Production	Releases Growth Factors	†Stem Cell Proliferation	↑ Mitochondrial Biogenesis
Platelet Activation PPE	1		1	$\sqrt{}$	\checkmark	
Nitric Oxide BDNF	V	√ √	√ √ √	√ √	$\sqrt{}$	\checkmark
CGRP CSD Serotonin	1	\ \ \	\ \ \ \	\ \ \	\ \ \	

Table 1.—Neuroprotective Properties of Migraine Components

released by blood platelets, increasing its plasma concentration. 134-136

Serotonin may contribute to migraine pain by producing inflammation and sensitizing sensory nerves. However, it is also involved in neuroprotection.

Serotonin has antioxidant properties in vitro¹³⁸ and an animal model of serotonin deficiency shows reduced antioxidant capacity. Moreover, serotonin promotes the differentiation of neurons and glia, and the formation of synapses. Acting through the 5-HT_{2B} receptor, serotonin facilitates neurogenesis in the dentate gyrus of the hippocampus. 140

In part, serotonin seems to exert these neurotrophic effects by releasing a number of growth factors. Thus, via the 5-HT_{1A} receptor, serotonin appears to trigger the release of a protein, S100B, from astrocytes. S100B resembles a neurotrophic factor in that it prevents apoptosis, facilitates the outgrowth of neurites, and modulates long-term synaptic plasticity.¹⁴¹ Moreover, serotonin causes astrocytes to raise their antioxidant support for neurons, by stimulating the release of metallothioneins, a class of antioxidant enzymes, and cysteine, which is taken up by neurons and used to produce the antioxidant glutathione.¹⁴¹

It appears that serotonin can also induce the production of BDNF, which as we have seen promotes neurogenesis, dendritic branching, synaptic plasticity, and neuronal survival (suppression of apoptosis) during ischemia, physical trauma, and psychosocial stress.^{89,91,92} Further, serotonin can stimulate

neurogenesis via insulin-like growth factor-1 (IGF-1)^{142,143} and limit glutamatergic neurotransmission through the release of VEGF and fractalkine. 144-147

Additional forms of neuroprotection might be suspected. Thus, in proximal tubular cells of the kidney, agonists at 5HT₂ receptors stimulate biogenesis of mitochondria, and thus presumably recovery from ischemic and other injuries. ¹⁴⁸

Through these mechanisms, serotonin protects neurons from ischemic and excitotoxic injury. All this may explain why blood levels of serotonin rise at the start of a migraine, as they do in closed head injury. In animal studies, 5-HT_{1A} agonists such as buspirone decrease lesion size and improve behavioral and cognitive recovery after experimental head injury or subdural hematoma, while in a clinical trial, fluoxetine, an SSRI and 5-HT_{1A} agonist, facilitated motor recovery following ischemic stroke, independently of effects on mood.

The effects of these migraine components are summarized in the Table 1. As may be seen, each component is neuroprotective through multiple mechanisms.

Interaction Among Components.—These components appear embedded in feedback loops, as shown schematically in Figure 1. For example, PAF stimulates the release of BDNF and serotonin from platelets and increases production of nitric oxide by the endothelium, which in turn raises production of BDNF. BDNF then furthers the production of nitric oxide in a positive feedback loop. CGRP and substance P may stimulate eNOS and increase

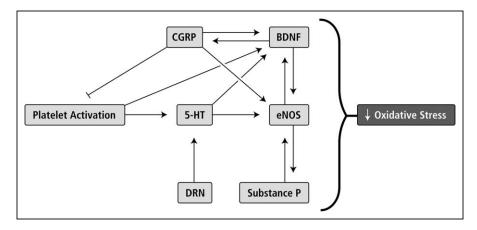


Fig. 1.—Interactions among migraine components. The components of a migraine attack are integrated by mostly positive, mutually reinforcing feedback loops. 5-HT = serotonin; BDNF = brain-derived neurotrophic factor; CGRP = calcitonin gene-related peptide; DRN = dorsal raphe nucleus; eNOS = endothelial nitric oxide synthase.

the production of nitric oxide. 154,155 Nitric oxide, in turn, may facilitate substance P release and, speculatively, CGRP neurotransmission in the trigeminal nucleus. 66,102 Serotonin, we have seen, stimulates endothelial nitric oxide production and activates a wide range of proliferative and prosurvival second messengers, including BDNF, VEGF, and IGF-1.

There are also indications of negative feedback loops, perhaps contributing to the selflimiting nature of migraine attacks. Thus, while PPE allows delivery of an antioxidant, albumin, to the CSF, in theory it could also allow entry of white blood cells, compromising the immune privilege of the CNS. However, at least in the mouse peritoneal cavity, CGRP attenuates the movement of neutrophils and monocytes through the endothelium, ¹⁵⁶ perhaps because CGRP inhibits the endothelial cells from producing certain chemokines as well as macrophage chemoattractant protein-1 and vascular cell adhesion molecule-1. 101,157 Thus, CGRP, an effector molecule for neurogenic inflammation, helps to maintain the non-immune nature of the inflammation. CGRP also limits platelet activation, 158 presumably helping to prevent excessive coagulation. Serotonin, meanwhile, through the release of fractalkine, 159,160 and directly through 5-HT_{2C} receptors, 161,162 limits the activation of microglia, helping to prevent neurogenic inflammation from spilling over into classical inflammation.

Thus, the suggestion is of an integrated system for neural protection and repair, coextensive with migraine.

DISCUSSION

Numerous components of the migraine attack—CSD, platelet activation, PPE, generation of nitric oxide, the release of CGRP, BDNF, substance P, and serotonin—and, we may surmise, the migraine itself—appear to be neuroprotective, reducing oxidant production and the energy requirement of neurons, stimulating neurogenesis, causing mitochondria to replicate, and delivering antioxidants and, especially, growth factors to the brain.

Nonetheless, the approach here—speculative, hypothesis-generating, preliminary—has a number of potential weaknesses:

First, the literature reviewed on the components of migraine was in vitro or on conditions other than migraine, primarily ischemia. The components may not have the same effect in the context of a migraine attack. However, ischemia may not be a far-fetched analogy, as migraines may be triggered by hypoxia, hypoglycemia, and presumably via CSD, by microembolisms, had all aspects of ischemia. Moreover, the fact that numerous components of a migraine effectively respond to a threat to the brain such as ischemia is consistent with the idea that primary migraine is also a response to threats to the brain.

Second, it must be considered that neurotrophic pathways are pleiotropic; the mere fact that they are activated does not mean that they will have a neuroprotective effect in migraine. We have seen, however, that at least in ischemia, the activation of these pathways does seem to be protective.

There are, in addition, arguments against *any* neuroprotective theory of migraines. Thus, high baseline attack frequency¹⁶⁵ and relative failure of acute medications¹⁶⁶ predict new-onset chronification, suggesting that migraines are progressive. Moreover, the frequency of migraine attacks and the duration of the disorder predict harm to the brain,¹⁶⁷ while in migraine with visual aura, the associated white matter lesions progress over time.¹⁶⁸ Further, migraine disorder increases the risk of conditions such as hypertension,¹⁶⁹ ischemic stroke,¹⁷⁰ myocardial infarction^{171,172} and, of note, Parkinson's disease.¹⁷³ How can such a broad swath of damage be reconciled with a neuroprotective process?

Relevant here is that neurotrophic signaling is bivalent. 174 That is, while BDNF, for example, is generally neuroprotective through the TrkB receptor, under certain circumstances BDNF can have the opposite effect. Thus, when neurons are subjected to severe mechanical stresses (as in traumatic brain injury), severe biochemical stresses (such as amyloid beta in Alzheimer's disease) or, apparently, severe psychosocial stress (as in major depressive disorder and posttraumatic stress disorder), they over-express a low affinity neurotrophin receptor, p75NTR, through which BDNF is proapoptotic and inhibits the proliferation of stem cells. 175 That is, high levels of physiological or psychosocial stress may connote an environment so harsh or damage so severe that apoptosis seems the best option. Under these conditions, a migraine attack could be neurodestructive. Similar to other stress-related disorders, processes that are adaptive under normal circumstances may become harmful under high allostatic load—environmental pressures to which adaptation exerts a toll on the individual. 176 Thus, migraines may resemble other defensive reactions such as fever, swelling, inflammation, and pain—promoting healing at moderate levels but causing damage when too intense.

This harm could occur through several other mechanisms. First, we have seen that CSD protects the brain during subsequent ischemia-reperfusion. However, when CSD follows traumatic brain injury, ischemic stroke, subarachnoid hemorrhage, or eliptiform activity, the combination of oligemia and high energy demands to restore ion homeostasis are thought to cause (further) ischemic damage. 177 These models differ from migraine in that they involve damaged tissue with impaired neurovascular coupling. In healthy tissue, as presumably characterizes migraine, CSD appears to be neuroprotective. Possibly, however, in small regions of cortex the vascular supply might be inadequate for the energy demands of CSD, causing localized ischemic damage. This might account for the progression of white matter lesions in migraine with aura.

Second, the combined presence of neurotrophic factors and nociceptive drive may lead to synaptic strengthening in pain pathways and conversion to a chronic disorder. As only 2.5% of migraineurs transition to the chronic state in a given year, this presumably describes only the most severe migraines.

Third, under some conditions migraines might increase oxidative stress in a vicious cycle. Thus, mast cells can raise the production of oxidants by the brain's renin-angiotensin system. 179,180 Activated platelets can produce oxidants by forming complexes with leukocytes. 181 Moreover, PPE is a means by which neurogenic inflammation (neuroprotective) can trigger classical inflammation, causing further oxidative damage. 182 We have seen that at moderate levels, albumin extravasated into cerebrospinal fluid likely functions as an antioxidant, facilitates repair of neuronal membranes, and causes astrocytes to release a trophic factor, oleic acid. However, albumin in CSF also may cause the proliferation of microglia¹⁸³ and promote inflammation. ¹⁸⁴ Naturalistically, a high concentration of albumin might indicate severe compromise of the blood-brain barrier and elicit a strong defensive response against pathogens.

Thus, the damage associated with migraine may be restricted to unusually intense, frequent, or prolonged attacks, migraines occurring in the context of severe physiological or psychosocial stress or, in the case of CSD, in regions of impaired vascular supply.

Alternatively, the damage may not be due to migraine attacks but to the migraine diathesis. The attacks may be simply a marker for the severity and persistence of the underlying diathesis. Consider, for example, the increased risk of hypertension in migraineurs. In animal models, CGRP, ¹⁸⁵ eNOS, ⁶⁷ and serotonin ¹⁸⁶ each protect against the development of hypertension. However, oxidative stress, a likely aspect of migraine vulnerability, is a risk factor for hypertension, ¹⁸⁷ endothelial dysfunction, ¹⁸⁸ and atherosclerosis. ¹⁸⁹

Thus, the contention here is that to truly solve migraines and the associated risk, we must identify and treat the underlying diathesis, rather than focusing on the body's defensive response to it.

Currently, CGRP is a promising therapeutic target in migraines. Thus, the evidence that CGRP may have a neuroprotective function raises the possibility, theoretical at this point, of long-term harm from anti-CGRP medications. 109,191

CONCLUSIONS

In all, then, there seems a fair amount of evidence that numerous components of a migraine attack, and presumably the attacks themselves, involve processes that protect and repair the brain. Of course, the existence of an integrated neuroprotective, neurorestorative system could be quite valuable, and would continue the grand tradition of migraines as a Rosetta stone, 192,193 illuminating normal brain functioning as well as disease, and informing the broad neurological field. For intrinsic to the human condition is a brain that must balance between sensation and excitotoxicity, between defense against pathogens and unintended selfdestruction, and between receptivity to the environment and exposure to its toxins. The migraine attack, as a particularly violent return to homeostasis, may have a key role in maintaining our neural integrity.

STATEMENT OF AUTHORSHIP

Category 1

(a) Conception and Design Jonathan M. Borkum

(b) Acquisition of Data

Jonathan M. Borkum

(c) Analysis and Interpretation of Data

Jonathan M. Borkum

Category 2

(a) Drafting the Manuscript

Jonathan M. Borkum

(b) Revising It for Intellectual Content

Jonathan M. Borkum

Category 3

(a) Final Approval of the Completed Manuscript

Jonathan M. Borkum

REFERENCES

- 1. Headache Classification Committee of the International Headache Society. The International Classification of Headache Disorders, 3rd edition (beta version). *Cephalalgia*. 2013;33:629-808.
- 2. Coppola G, Pierelli F, Schoenen J. Is the cerebral cortex hyperexcitable or hyperresponsive in migraine? *Cephalalgia*. 2007;27:1427-1439.
- 3. Welch KM, D'Andrea G, Tepley N, Barkeley G, Ramadan NM. The concept of migraine as a state of central neuronal hyperexcitability. *Neurol Clin.* 1990;8:817-828.
- Goadsby PJ, Lipton RB, Ferrari MD. Migraine current understanding and treatment. N Engl J Med. 2002;346:257-270.
- Lance JW, Lambert GA, Goadsby PJ, Duckworth JW. Brain stem influences on the cephalic circulation: Experimental data from cat and monkey of relevance to the mechanism of migraine. *Headache*. 1983;23:258-265.
- 6. Weiller C, May A, Limmroth V, et al. Brain stem activation in spontaneous human migraine attacks. *Nat Med.* 1995;1:658-660.
- 7. Lambert GA, Hoskin KL, Zagami AS. Cortico-NRM influences on trigeminovascular sensation. *Cephalalgia*. 2008;28:640-652.
- 8. Loder E. What is the evolutionary advantage of migraine? *Cephalalgia*. 2002;22:624-632.
- 9. Goadsby P. What is the physiological role of the trigeminovascular system? *Cephalalgia*. 1995;15: 333.
- 10. Cortelli P, Montagna P. Migraine as visceral pain. *Neurol Sci.* 2009;30:s19-s22.

- 11. Eren Y, Dirik E, Neşelioğlu S, Erel Ö. Oxidative stress and decreased thiol level in patients with migraine: Cross-sectional study. *Acta Neurol Belg.* 2015;115:643-649.
- 12. Geyik S, Altunisik E, Neyal AM, Taysi S. Oxidative stress and DNA damage in patients with migraine. *J Headache Pain*. 2016;17:10.
- 13. Neri M, Frustaci A, Milic M, et al. A metaanalysis of biomarkers related to oxidative stress and nitric oxide pathway in migraine. *Cephalalgia.* 2015;35:931-937.
- 14. Shatillo A, Koroleva K, Giniatullina R, et al. Cortical spreading depression induces oxidative stress in the trigeminal nociceptive system. *Neuroscience*. 2013;253:341-349.
- 15. Aytaç B, Coşkun Ö, Alioğlu B, et al. Decreased antioxidant status in migraine patients with brain white matter hyperintensities. *Neurol Sci.* 2014; 35:1925-1929.
- 16. Ripa P, Ornello R, Pistoia F, Carolei A, Sacco S. The renin-angiotensin system: A possible contributor to migraine pathogenesis and prophylaxis. *Expert Rev Neurother.* 2014;14:1043-1055.
- 17. Paterna S, Di Pasquale P, D'Angelo A, et al. Angiotensin-converting enzyme gene deletion polymorphism determines an increase in frequency of migraine attacks in patients suffering from migraine without aura. *Eur Neurol.* 2000;43: 133-136.
- 18. MacClellan LR, Howard TD, Cole JW, et al. Relation of candidate genes that encode for endothelial function to migraine and stroke: The Stroke Prevention in Young Women study. *Stroke*. 2009;40:e550-e557.
- 19. Geyik S, Ergun S, Kuzudişli S, et al. Plasma urotensin-2 level and Thr21Met but not Ser89Asn polymorphisms of the urotensin-2 gene are associated with migraines. *J Headache Pain*. 2016;17: 36.
- 20. Borkum JM. Migraine triggers and oxidative stress: A narrative review and synthesis. *Headache*. 2016;56:12-35.
- 21. Dong X, Guan X, Chen K, et al. Abnormal mitochondrial dynamics and impaired mitochondrial biogenesis in trigeminal ganglion neurons in a rat model of migraine. *Neurosci Lett.* 2017;636:127-133.
- 22. Markley HG. Coenzyme Q10 and riboflavin: The mitochondrial connection. *Headache*. 2012;52:81-87.

- 23. Barbiroli B, Montagna P, Cortelli P, et al. Abnormal brain and muscle energy metabolism show by ³¹P magnetic resonance spectroscopy in patients affected by migraine with aura. *Neurology*. 1992;42:1209-1214.
- 24. Reyngoudt H, Achten E, Paemeleire K. Magnetic resonance spectroscopy in migraine: What have we learned so far? *Cephalalgia*. 2012;32:845-859.
- 25. Welch KM, Levine SR, D'Andrea G, Schultz LR, Helpern JA. Preliminary observations on brain energy metabolism in migraine studied by in vivo phosphorous 31 NMR spectroscopy. *Neurology*. 1989;39:538-541.
- 26. Stuart S, Griffiths LR. A possible role for mitochondrial dysfunction in migraine. *Mol Genet Genomics*. 2012;287:837-844.
- 27. Wu Y-T, Wu S-B, Wei Y-H. Roles of sirtuins in the regulation of antioxidant defense and bioenergetic function of mitochondria under oxidative stress. *Free Radic Res.* 2014;48:1070-1084.
- 28. Huang D, Li S, Dhaka A, Story GM, Cao Y-Q. Expression of the transient receptor potential channels TRPV1, TRPA1 and TRPM8 in mouse trigeminal primary afferent neurons innervating the dura. *Mol Pain*. 2012:8:66.
- 29. Kozai D, Ogawa N, Mori Y. Redox regulation of transient receptor potential channels. *Antioxid Redox Signal*. 2014;21:971-986.
- 30. Benemei S, Fusi C, Trevisan G, Geppetti P. The TRPA1 channel in migraine mechanism and treatment. *Br J Pharmacol*. 2014;171:2552-2567.
- Dussor G, Yan J, Xie JY, Ossipov MH, Dodick DW, Porreca F. Targeting TRP channels for novel migraine therapeutics. ACS Chem Neurosci. 2014;5:1085-1096.
- 32. Schiavone S, Jaquet V, Trabace L, Krause K-H. Severe life stress and oxidative stress in the brain: From animal models to human pathology. *Antioxid Redox Signal*. 2013;18:1475-1490.
- Abdel-Salam OME, Salem NA, Hussein JS. Effect of aspartame on oxidative stress and monoamine neurotransmitter levels in lipopolysaccharide-treated mice. Neurotox Res. 2012;21:245-255.
- 34. Cahill-Smith S, Li JM. Oxidative stress, redox signaling and endothelial dysfunction in ageing-related neurodegenerative diseases: A role of NADPH oxidase 2. *Br J Clin Pharmacol*. 2014; 78:441-453.

- 35. Goadsby PJ. Pathophysiology of migraine. *Ann Indian Acad Neurol.* 2012;15:s15-s22.
- Goadsby PJ, Edvinsson L, Ekman R. Vasoactive peptide release in the extracerebral circulation of humans during migraine headache. *Ann Neurol*. 1990;28:183-187.
- 37. Olesen J, Burstein R, Ashina M, Tfelt-Hansen P. Origin of pain in migraine: Evidence for peripheral sensitization. *Lancet Neurol.* 2009;8:679-690.
- 38. Ho TW, Edvinsson L, Goadsby PJ. CGRP and its receptors provide new insights into migraine pathophysiology. *Nat Rev Neurol.* 2010;6:573-582.
- 39. Burstein R, Noseda R, Borsook D. Migraine: Multiple processes, complex pathophysiology. *J Neurosci.* 2015;35:6619-6629.
- 40. Banoujaafar H, Monnier A, Pernet N, et al. Brain BDNF levels are dependent on cerebrovascular endothelium-derived nitric oxide. Eur J Neurosci. 2016;44:2226-2235.
- 41. Serra-Millàs M. Are the changes in the peripheral brain-derived neurotrophic factor levels due to platelet activation? *World J Psychiatr.* 2016;6:84-101.
- 42. Chu CT, Levinthal DJ, Kulich SM, Chalovich EM, DeFranco DB. Oxidative neuronal injury: The dark side of ERK1/2. *Eur J Biochem.* 2004; 271:2060-2066.
- 43. Moosavi F, Hosseini R, Saso L, Firuzi O. Modulation of neurotrophic signaling pathways by polyphenols. *Drug Des Devel Ther.* 2016;10:23-42.
- 44. Patapoutian A, Reichardt LF. Trk receptors: Mediators of neurotrophin action. *Curr Opin Neurobiol.* 2001;11:272-280.
- 45. Danese E, Montagnana M, Lippi G. Platelets and migraine. *Thromb Res.* 2014;134:17-22.
- 46. Hanington E. Migraine: The platelet hypothesis after ten years. *Biomed Pharmacother*. 1989;43: 719-726.
- 47. Hanington E, Jones RJ, Amess JA, Wachowicz B. Migraine: A platelet disorder. *Lancet.* 1981; 318:720-723.
- 48. Sarchielli P, Alberti A, Coppola F, et al. Platelet-activating factor (PAF) in internal jugular venous blood of migraine without aura patients assessed during migraine attacks. *Cephalalgia*. 2004;24:623-630.
- 49. Burnouf T, Goubran HA, Chou M-L, Devos D, Radosevic M. Platelet microparticles: Detection and assessment of their paradoxical functional

- roles in disease and regenerative medicine. *Blood Rev.* 2014;28:155-166.
- 50. Hayon Y, Dashevsky O, Shai E, Varon D, Leker RR. Platelet microparticles promote neural stem cell proliferation, survival, and differentiation. *J Mol Neurosci.* 2012;47:659-665.
- 51. Hayon Y, Dashevsky O, Shai E, Brill A, Varon D, Leker RR. Platelet microparticles induce angiogenesis and neurogenesis after cerebral ischemia. *Curr Neurovasc Res.* 2012;9:185-192.
- 52. Hayon Y, Shai E, Varon D, Leker RR. The role of platelets and their microparticles in rehabilitation of ischemic brain tissue. *CNS Neurol Disord Drug Targets*. 2012;11:921-925.
- 53. Paparone S, Severini C, Ciotti MT, D'Agata V, Calissano P, Cavallaro S. Transcriptional landscapes at the intersection of neuronal apoptosis and substance P-induced survival: Exploring pathways and drug targets. *Cell Death Discov*. 2016;2: 16050.
- 54. Taverna M, Marie A-L, Mira J-P, Guidet B. Specific antioxidant properties of human serum albumin. *Ann Intensive Care*. 2013;3:4.
- Prajapati KD, Sharma SS, Roy N. Current perspectives on potential role of albumin in neuroprotection. *Rev Neurosci.* 2011;22:355-363.
- 56. Remmers M, Schmidt-Kastner R, Belayev L, Lin B, Busto R, Ginsberg MD. Protein extravasation and cellular uptake after high-dose human-albumin treatment of transient focal cerebral ischemia in rats. *Brain Res.* 1999;827:237-242.
- 57. Belayev L, Liu Y, Zhao W, Busto R, Ginsberg MD. Human albumin therapy of acute ischemic stroke: Marked neuroprotective efficacy at moderate doses and with a broad therapeutic window. *Stroke*. 2001;32:553-560.
- 58. Rodriguez de Turco EB, Belayev L, Liu Y, et al. Systemic fatty acid responses to transient focal cerebral ischemia: Influence of neuroprotectant therapy with human albumin. *J Neurochem.* 2002; 83:515-524.
- 59. Bento-Abreu A, Velasco A, Polo-Hernandez E, Perez-Reyes PL, Tabernero A, Medina JM. Megalin is a receptor for albumin in astrocytes and is required for the synthesis of the neurotrophic factor oleic acid. *J Neurochem*. 2008;106: 1149-1159.
- Tabernero A, Velasco A, Granda B, Lavado EM, Medina JM. Transcytosis of albumin in astrocytes

activates the sterol regulatory element-binding protein-1, which promotes the synthesis of the neurotrophic factor oleic acid. *J Biol Chem.* 2002; 277:4240-4246.

- 61. Ginsberg MD, Palesch YY, Hill MD, et al. High-dose albumin treatment for acute ischemic stroke (ALIAS) Part 2: A randomized, double-blind, phase 3, placebo-controlled trial. *Lancet Neurol*. 2013;12:1049-1058.
- 62. Hill MD, Martin RH, Palesch YY, et al. Albumin administration in acute ischemic stroke: Safety analysis of the ALIAS Part 2 multicenter trial. *PLoS One.* 2015;10:e0131390.
- 63. Suarez JI, Martin RH, Calvillo E, Bershad EM, Venkatasubba Rao CP. Effect of human albumin on TCD vasospasm, DCI, and cerebral infarction in subarachnoid hemorrhage: The ALISAH study. *Acta Neurochir Suppl.* 2015;120:287-290.
- 64. Sarchielli P, Alberti A, Codini M, Floridi A, Gallai V. Nitric oxide metabolites, prostaglandins and trigeminal vasoactive peptides in internal jugular vein blood during spontaneous migraine attacks. *Cephalalgia*. 2000;20:907-918.
- 65. Manivet P, Mouillet-Richard S, Callebert J, et al. PDZ-dependent activation of nitric oxide synthases by the serotonin 2B receptor. *J Biol Chem.* 2000;275:9324-9331.
- 66. Yonehara N, Yoshimura M. Effect of nitric oxide on substance P release from the peripheral endings of primary afferent neurons. *Neurosci Lett.* 1999;271:199-201.
- 67. González J, Valls N, Brito R, Rodrigo R. Essential hypertension and oxidative stress: New insights. *World J Cardiol.* 2014;6:353-366.
- 68. Liu H, Li J, Zhao F, Wang H, Qu Y, Mu D. Nitric oxide synthase in hypoxic or ischemic brain injury. *Rev Neurosci.* 2015;26:105-117.
- 69. Huang PL. Mouse models of nitric oxide synthase deficiency. *J Am Soc Nephrol.* 2000;11: s120-s123.
- Astort F, Mercau M, Giordanino E, et al. Nitric oxide sets off an antioxidant response in adrenal cells: Involvement of sGC and Nrf2 in HO-1 induction. *Nitric Oxide*. 2014;37:1-10.
- 71. Essler S, Dehne N, Brune B. Role of sestrin2 in peroxide signaling in macrophages. *FEBS Lett.* 2009;583:3531-3535.
- 72. Kurauchi Y, Hisatsune A, Isohama Y, Sawa T, Akaike T, Katsuki H. Nitric oxide/soluble

- guanylyl cyclase signaling mediates depolarization-induced protection of rat mesence-phalic dopaminergic neurons from MPP(+) cytotoxicity. *Neuroscience*. 2013;231:206-215.
- 73. Nisoli E, Tonello C, Cardile A, et al. Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. *Science*. 2005; 310:314-317.
- 74. Asada M, Ebihara S, Yamanda A, et al. Depletion of serotonin and selective inhibition of 2B receptor suppressed tumor angiogenesis by inhibiting endothelial nitric oxide synthase and extracellular signal-regulated kinase 1/2 phosphorylation. *Neoplasia*. 2009;11:408-417.
- Chen J, Zacharek A, Zhang C, et al. Endothelial nitric oxide synthase regulates brain-derived neurotrophic factor expression and neurogenesis after stroke in mice. *J Neurosci.* 2005;25:2366-2375.
- Haul S, Godecke A, Schrader J, Haas HL, Luhmann HJ. Impairment of neocortical longterm potentiation in mice deficient of endothelial nitric oxide synthase. *J Neurophysiol*. 1999;81: 494-497.
- 77. Monnier A, Prigent-Tessier A, Quirié A, et al. Brain-derived neurotrophic factor of the cerebral microvasculature: A forgotten and nitric oxidedependent contributor of brain-derived neurotrophic factor in the brain. *Acta Physiol.* 2017;219: 790-802.
- Guo S, Kim WJ, Lok J, et al. Neuroprotection via matrix-trophic coupling between cerebral endothelial cells and neurons. *Proc Natl Acad Sci USA*. 2008;105:7582-7587.
- Meuchel IW, Thompson MA, Cassivi SD, Pabelick CM, Prakash YS. Neurotrophins induce nitric oxide generation in human pulmonary artery endothelial cells. *Cardiovasc Res.* 2011;91: 668-676.
- 80. Lipsky RH, Marini AM. Brain-derived neurotrophic factor in neuronal survival and behavior-related plasticity. *Ann N Y Acad Sci.* 2007;1122:130-143.
- 81. Tanure MT, Gomez RS, Hurtado RC, Teixeira AL, Dominguez RB. Increased serum levels of brain-derived neurotrophic factor during migraine attacks: A pilot study. *J Headache Pain*. 2010;11: 427-430.
- 82. Blandini F, Rinaldi L, Tassorelli C, et al. Peripheral levels of BDNF and NGF in primary headaches. *Cephalalgia*. 2006;26:136-142.

- 83. Tietjen GE, Khubchandani J. Platelet dysfunction and stroke in the female migraineurs. *Curr Pain Headache Rep.* 2009;13:386-391.
- 84. Urbach A, Bruehl C, Witte OW. Microarray-based long-term detection of genes differentially expressed after cortical spreading depression. *Eur J Neurosci.* 2006;24:841-856.
- 85. Buldyrev I, Tanner NM, Hsieh HY, Dodd EG, Nguyen LT, Balkowiec A. Calcitonin generelated peptide enhances release of native brainderived neurotrophic factor from trigeminal ganglion neurons. *J Neurochem.* 2006;99:1338-1350.
- 86. Pezet S, McMahon SB. Neurotrophins: Mediators and modulators of pain. *Annu Rev Neurosci.* 2006;29:507-538.
- 87. Bowling H, Bhattacharya A, Klann E, Chao MV. Deconstructing brain-derived neurotrophic factor actions in adult brain circuits to bridge and existing informational gap in neuro-cell biology. *Neural Regen Res.* 2016;11:363-367.
- 88. Gonzalez A, Moya-Alvarado G, Gonzalez-Billaut C, Bronfman FC. Cellular and molecular mechanisms regulating neuronal growth by brainderived neurotrophic factor. *Cytoskeleton*. 2016; 73:612-628.
- 89. Binder DK, Scharfman HE. Brain-derived neurotrophic factor. *Growth Factors.* 2004;22:123-131.
- 90. Kiprianova I, Sandkuhler J, Schwab S, Hoyer S, Spranger M. Brain-derived neurotrophic factor improves long-term potentiation and cognitive functions after transient forebrain ischemia in the rat. *Exp Neurol.* 1999;159:511-519.
- 91. Larsson E, Nanobashvili A, Kokaia Z, Lindvall O. Evidence for neuroprotective effects of endogenous brain-derived neurotrophic factor after global forebrain ischemia in rats. *J Cereb Blood Flow Metab.* 1999;19:1220-1228.
- 92. Mattson MP, Maudsley S, Martin B. BDNF and 5-HT: A dynamic duo in age-related neuronal plasticity and neurodegenerative disorders. *Trends Neurosci.* 2004;27:589-594.
- 93. Radecki DT, Brown LM, Martinez J, Teyler TJ. BDNF protects against stress-induced impairments in spatial learning and memory and LTP. *Hippocampus*. 2005;15:246-253.
- 94. Foltran RB, Diaz SL. BDNF isoforms: A round trip ticket between neurogenesis and serotonin? *J Neurochem.* 2016;138:204-221.

- 95. Bac SH, Sung SH, Oh SY, et al. Sestrins activate Nrf2 by promoting p62-dependent autophagic degradation of Keap1 and prevent oxidative liver damage. *Cell Metab.* 2013;17:73-84.
- 96. Wu C-L, Chen S-D, Yin J-H, Hwang C-S, Yang D-I. Nuclear factor-kappaB-dependent sestrin2 induction mediates the antioxidant effects of BDNF against mitochondrial inhibition in rat cortical neurons. *Mol Neurobiol*. 2016;53:4126-4142.
- 97. Zoladz JA, Majerczak J, Zeligowska E, et al. Moderate-intensity interval training increases serum brain-derived neurotrophic factor level and decreases inflammation in Parkinson's disease patients. *J Physiol Pharmacol.* 2014;65:441-448.
- 98. Wild EJ, Tabrizi SJ. Targets for future clinical trials in Huntington's disease: What's in the pipeline? *Mov Disord.* 2014;29:1434-1445.
- 99. Ploughman M, Austin MW, Glynn L, Corbett D. The effects of poststroke aerobic exercise on neuroplasticity: A systematic review of animal and clinical studies. *Transl Stroke Res.* 2015;6:13-28.
- 100. Karsan N, Goadsby PJ. Calcitonin gene-related peptide and migraine. *Curr Opin Neurol.* 2015;28: 250-254.
- Russell FA, King R, Smillie S-J, Kodji X, Brain SD. Calcitonin gene-related peptide: Physiology and pathophysiology. *Physiol Rev.* 2014;94:1099-1142.
- 102. Messlinger K, Lennerz JK, Eberhardt M, Fischer MJM. CGRP and NO in the trigeminal system: Mechanisms and role in headache generation. *Headache*. 2012;52:1411-1427.
- 103. Fei S, Wei S, Jie-Ming M, Xian W. Calcitonin gene-related peptide therapy suppresses reactive oxygen species in the pancreas and prevents mice from autoimmune diabetes. *Acta Physiol Sin.* 2003;55:625-632.
- 104. Zhou Z, Hu C-P, Wang C-J, Li T-T, Peng J, Li Y-J. Calcitonin gene-related peptide inhibits angiotensin II-induced endothelial progenitor cells senescence through up-regulation of klotho expression. *Atherosclerosis*. 2010;213:92-101.
- 105. Schaeffer C, Vandroux D, Thomassin L, Athias P, Rochette L, Connat J-L. Calcitonin generelated peptide partly protects cultured smooth muscle cells from apoptosis induced by an oxidative stress via activation of ERK1/2 MAPK. *Biochim Biophys Acta*. 2003;1643:65-73.

- 106. Ringer C, Tune S, Bertoune MA, et al. Disruption of calcitonin gene-related peptide signaling accelerates muscle denervation and dampens cytotoxic neuroinflammation in SOD1 mutant mice. *Cell Mol Life Sci.* 2017;74:339-358.
- 107. Reddington M, Priller J, Treichel J, et al. Astrocytes and microglia as potential targets for calcitonin gene related peptide in the central nervous system. *Can J Physiol Pharmacol*. 1995;73:1047-1049.
- 108. Zhang J-Y, Yan G-T, Liao J, et al. Leptin attenuates cerebral ischemia/reperfusion injury partially by CGRP expression. *Eur J Pharmacol*. 2011;671: 61-69.
- 109. Abushik PA, Bart G, Korhonen P, et al. Pronociceptive migraine mediator CGRP provides neuroprotection of sensory, cortical and cerebellar neurons via multi-kinase signaling. *Cephalalgia*. In press; DOI: 10.1177/0333102416681588
- 110. Bulloch K, Prasad A, Conrad CD, McEwen BS, Milner TA. Calcitonin gene-related peptide level in the rat dentate gyrus increases after damage. *Neuroreport.* 1996;7:1036-1040.
- 111. Bulloch K, Milner TA, Prasad A, Hsu M, Buzsaki G, McEwen BS. Induction of calcitonin gene-related peptide-like immunoreactivity in hippocampal neurons following ischemia: A putative regional modulator of the CNS injury/immune response. *Exp Neurol.* 1998;150:195-205.
- 112. Edvinsson L, Olesen IJ, Kingman TA, McCulloch J, Uddman R. Modification of vasoconstrictor responses in cerebral blood vessels by lesioning of the trigeminal nerve: Possible involvement of CGRP. *Cephalalgia*. 1995;15:373-383.
- 113. Gherardini G, Evans GR, Theodorsson E, et al. Calcitonin gene-related peptide in experimental ischemia. Implication of an endogenous anti-ischemic effect. *Ann Plast Surg.* 1996;36:616-620.
- 114. Kovács A, Telegdy G. Effect of intracerebroventricular administration of calcitonin gene-related peptide on passive avoidance behavior in rats. *Neuropeptides.* 1992;23:51-54.
- 115. Hashikawa-Hobara N, Ogawa T, Sakamoto Y, et al. Calcitonin gene-related peptide pre-administration acts as a novel antidepressant in stressed mice. *Sci Rep.* 2015;5:12559.
- 116. Johnston FG, Bell BA, Robertson IJA, et al. Effect of calcitonin-gene-related peptide

- postoperative neurological deficits after subarachnoid haemorrhage. *Lancet*. 1990;335:869-872.
- 117. European CGRP in Subarachnoid Haemorrhage Study Group. Effect of calcitonin-gene-related peptide in patients with delayed postoperative cerebral ischaemia after aneurysmal subarachnoid haemorrhage. *Lancet.* 1992;339:831-834.
- 118. Zhao Y-Z, Lin M, Lin Q, et al. Intranasal delivery of bFGF with nanoliposomes enhances in vivo neuroprotection and neural injury recovery in a rodent stroke model. *J Control Release*. 2016; 224:165-175.
- 119. Cui Y, Kataoka Y, Watanabe Y. Role of cortical spreading depression in the pathophysiology of migraine. *Neurosci Bull.* 2014;30:812-822.
- 120. Bolay H, Reuter U, Dunn AK, Huang Z, Boas DA, Moskowitz MA. Intrinsic brain activity triggers trigeminal meningeal afferents in a migraine model. *Nat Med.* 2002;8:136-142.
- 121. Marini AM, Popolo M, Pan H, Blondeau N, Lipsky RH. Brain adaptation to stressful stimuli: A new perspective on potential therapeutic approaches based on BDNF and NMDA receptors. *CNS Neurol Disord Drug Targets*. 2008;7: 382-390.
- 122. Viggiano E, Viggiano D, Viggiano A, De Luca B, Monda M. Cortical spreading depression increases the phosphorylation of AMP-activated protein kinase in the cerebral cortex. *Neurochem Res.* 2014;39:2431-2439.
- 123. Choudhuri R, Cui L, Yong C, et al. Cortical spreading depression and gene regulation: Relevance to migraine. *Ann Neurol.* 2002;51:499-506.
- 124. Viggiano E, Monda V, Messina A, et al. cortical spreading depression produces a neuroprotective effect activating mitochondrial uncoupling protein-5. *Neuropsychiatr Dis Treat.* 2016;12:1705-1710.
- 125. Kariko K, Harris VA, Rangel Y, Duvall ME, Welsh FA. Effect of cortical spreading depression on the levels of mRNA coding for putative neuroprotective proteins in rat brain. *J Cereb Blood Flow Metab.* 1998;18:1308-1315.
- 126. Kawahara N, Croll SD, Wiegand SJ, Klatzo I. Cortical spreading depression induces long-term alterations of BDNF levels in cortex and hippocampus distinct from lesion effects: Implications for ischemic tolerance. *Neurosci Res.* 1997;29:37-47.

- 127. Rangel YM, Kariko K, Harris VA, Duvall ME, Welsh FA. Dose-dependent induction of mRNAs encoding brain-derived neurotrophic factor and heat-shock protein-72 after cortical spreading depression in the rat. *Brain Res Mol Brain Res*. 2001;88:103-112.
- 128. Faraguna U, Nelson A, Vyazovskiy VV, Cirelli C, Tononi G. Unilateral cortical spreading depression affects sleep need and induces molecular and electrophysiological signs of synaptic potentiation in vivo. *Cereb Cortex*. 2010;20:2939-2947.
- 129. Hamel E. Serotonin and migraine: Biology and clinical implications. *Cephalalgia*. 2007;27:1293-1300.
- 130. Sakai Y, Dobson C, Diksic M, Aubé M, Hamel E. Sumatriptan normalizes the migraine attackrelated increase in brain serotonin synthesis. *Neurology*. 2008;70:431-439.
- 131. Raskin NH, Hosobuchi Y, Lamb S. Headache may arise from perturbation of brain. *Headache*. 1987;27:416-420.
- 132. Azmitia EC. Serotonin neurons, neuroplasticity, and homeostasis of neural tissue. *Neuropsychopharmacology*. 1999;21:s33-s45.
- 133. Azmitia EC. Serotonin and brain: Evolution, neuroplasticity, and homeostasis. *Int Rev Neurobiol.* 2007;77:31-56.
- 134. Ferrari MD, Odink J, Tapparelli C, Van Kempen GMJ, Pennings EJM, Bruyn GW. Serotonin metabolism in migraine. *Neurology*. 1989;39:1239-1242.
- 135. Izzati-Zade KF. The role of serotonin in the pathogenesis and clinical presentations of migraine attacks. *Neurosci Behav Physiol.* 2008;38:501-505.
- 136. Sicuteri F, Testi A, Anselmi B. Biochemical investigations in headache: Increase in hydroxyindoleacetic acid excretion during migraine attacks. *Int Arch Allergy.* 1961;19:55-58.
- 137. Lambert GA. The lack of peripheral pathology in migraine headache. *Headache*. 2010;50:895-908.
- 138. Kalogiannis M, Delikatny EJ, Jeitner TM. Serotonin as a putative scavenger of hypohalous acid in the brain. *Biochim Biophys Acta*. 2016;1862: 651-661.
- 139. Weng R, Shen S, Tian Y, et al. Metabolomics approach reveals integrated metabolic network associated with serotonin deficiency. *Sci Rep.* 2015;5:11864.

- 140. Diaz SL, Doly S, Narboux-Nême N, et al. 5-HT(2b) receptors are required for serotonin-selective antidepressant actions. *Mol Psychiatry*. 2012;17:154-163.
- 141. Miyazaki I, Asanuma M. Serotonin 1A receptors on astrocytes as a potential target for the treatment of Parkinson's disease. *Curr Med Chem.* 2016;23:686-700.
- 142. Lauder JM. Neurotransmitters as growth regulatory signals: Role of receptors and second messengers. *Trends Neurosci.* 1993;16:233-240.
- 143. Aberg MA, Aberg ND, Palmer TD, et al. IGF-1 has a direct proliferative effect in adult hippocampal progenitor cells. *Mol Cell Neurosci.* 2003; 24:23-40.
- 144. Greene J, Banasr M, Lee B, Warner-Schmidt J, Duman RS. Vascular endothelial growth factor signaling is required for the behavioral actions of antidepressant treatment: Pharmacological and cellular characterization. *Neuropsychopharmacology*. 2009;34:2459-2468.
- 145. Scharfman HE, MacLusky NJ. Estrogen-growth factor interactions and their contributions to neurological disorders. *Headache*. 2008;48:s77-s89.
- 146. Heinisch S, Kirby LG. Fractalkine/CX3CL1 enhances GABA synaptic activity at serotonin neurons in the rat dorsal raphe nucleus. *Neuroscience*. 2009;164:1210-1223.
- 147. Ragozzino D, Di Angelantonio S, Trettel F, et al. Chemokine fractalkine/CX₃CL1 negatively modulates active glutamatergic synapses in rat hippocampal neurons. *J Neurosci.* 2006;26:10488-10498.
- 148. Rasbach KA, Funk JA, Jayavelu T, Green PT, Schnellmann RG. 5-hydroxytryptamine receptor stimulation of mitochondrial biogenesis. *J Pharmacol Exp Ther.* 2010;332:632-639.
- 149. Semkova I, Wolz P, Krieglstein J. Neuroprotective effect of 5 HT_{1A}-receptor agonist, Bay x 3702, demonstrated in vitro and in vivo. *Eur J Pharmacol.* 1998;359:251-260.
- 150. Sharma HS, Patnaik R, Patnaik S, Mohanty S, Sharma A, Vannemreddy P. Antibodies to serotonin attenuate closed head injury induced bloodbrain barrier disruption and brain pathology. *Ann N Y Acad Sci.* 2007;1122:295-312.
- 151. Cheng JP, Leary JB, Sembhi A, Edwards CM, Bondi CO, Kline AE. 5-hydroxytriptamine_{1A} (5-HT_{1A}) receptor agonists: A decade of empirical evidence supports their use as an efficacious

- therapeutic strategy for brain trauma. *Brain Res.* 2016:1640:5-14.
- 152. Chollet F, Tardy J, Albucher J-F, et al. Fluoxetine for motor recovery after acute ischaemic stroke (FLAME): A randomised placebocontrolled trial. *Lancet Neurol.* 2011;10:123-130.
- 153. Duran WN, Seyama A, Yoshimura K, et al. Stimulation of NO production and of eNOS phosphorylation in the microcirculation in vivo. *Microvasc Res.* 2000;60:104-111.
- 154. Kokkoris S, Andrews P, Webb DJ. Role of calcitonin gene-related peptide in cerebral vasospasm, and as a therapeutic approach to subarachnoid hemorrhage. *Front Endocrinol.* 2012;3:135.
- 155. Yonehara N, Yoshimura M. Interaction between nitric oxide and substance P on heat-induced inflammation in rat paw. *Neurosci Res.* 2000;36:35-43.
- 156. Gomes RN, Castro-Faria-Neto HC, Bozza PT, et al. Calcitonin gene-related peptide inhibits local acute inflammation and protects mice against lethal endotoxemia. *Shock.* 2005;24:590-594.
- 157. Huang J, Stohl LL, Zhou X, Ding W, Granstein RD. Calcitonin gene-related peptide inhibits chemokine production by human dermal microvascular endothelial cells. *Brain Behav Immun*. 2011; 25:787-799.
- 158. Matsumoto Y, Ueda S, Matsushita S, Ozawa T, Yamaguchi H. Calcitonin gene-related peptide inhibits human platelet aggregation. *Jpn Circ J*. 1996;60:797-804.
- 159. Cardona AE, Pioro EP, Sasse ME, et al. Control of microglial neurotoxicity by the fractalkine receptor. *Nat Neurosci.* 2006;9:917-924.
- 160. Corona AW, Huang Y, O'Connor JC, et al. Fractalkine receptor (CX3CR1) deficiency sensitizes mice to the behavioral changes induced by lipopolysaccharide. *J Neuroinflammation*. 2010;7:93.
- 161. Abdel-Salam OM, Baiuomy AR, Arbid MS. Studies on the anti-inflammatory effect of fluoxetine in the rat. *Pharmacol Res.* 2004;49:119-131.
- Hwang J, Zheng LT, Ock J, Lee MG, Suk K. Antiinflammatory effects of m-chlorophenylpiperazine in brain glia cells. *Int Immunopharmacol*. 2008;8:1686-1694.
- 163. Nozari A, Dilekoz E, Sukhotinsky I, et al. Microemboli may link spreading depression, migraine aura, and patent foramen ovale. *Ann Neurol*. 2010;67:221-229.

- 164. Caputi L, Usai S, Carriero MR, et al. Microembolic air load during contrast-transcranial Doppler: A trigger for migraine with aura? *Headache*. 2010;50:1320-1327.
- 165. Lipton RB. Tracing transformation: Chronic migraine classification, progression, and epidemiology. *Neurology*. 2009;72:s3-s7.
- 166. Lipton RB, Fanning KM, Serrano D, Reed ML, Cady R, Buse DC. Ineffective acute treatment of episodikc migraine is associated with newonset chronic migraine. *Neurology*. 2015;84:688-695.
- 167. Schmitz N, Admiraal-Behloul F, Arkink EB, et al. Attack frequency and disease duration as indicators for brain damage in migraine. *Head-ache*. 2008;48:1044-1055.
- 168. Dinia L, Bonzano L, Albano B, et al. White matter lesions progression in migraine with aura: A clinical and MRI longitudinal study. *J Neuroimaging*. 2013; 23:47-52.
- 169. Entonen AH, Suominen SB, Korkeila K, et al. Migraine predicts hypertension a cohort study of the Finnish working-age population. *Eur J Public Health*. 2013;24:244-248.
- 170. Kurth T. The association of migraine with ischemic stroke. *Curr Neurol Neurosci Rep.* 2010;10: 133-139.
- 171. Kurth T, Gaziano JM, Cook NR, et al. Migraine and risk of cardiovascular disease in men. *Arch Intern Med.* 2007;167:795-801.
- 172. Kurth T, Gaziano JM, Cook NR, Logroscino G, Diener HC, Buring JE. Migraine and risk of cardiovascular disease in women. *JAMA*. 2006;296: 283-291.
- 173. Wang H-I, Ho Y-C, Huang Y-P, Pan S-L. Migraine is related to an increased risk of Parkinson's disease: A population-based, propensity score-matched, longitudinal follow-up study. *Cephalalgia*. 2016;36:1316-1323.
- 174. Wu Y, Liu W-Z, Liu T, Feng X, Yang N, Zhou H-F. Signaling pathway of MAPK/ERK in cell proliferation, differentiation, migration, senescence and apoptosis. *J Recept Signal Transduct*. 2015;35:600-604.
- 175. Shu Y-H, Lu X-M, Wei J-X, Xiao L, Wang Y-T. Update on the role of p75NTR in neurological disorders: A novel therapeutic target. *Biomed Pharmacother*. 2015;76:17-23.

- 176. Borsook D, Maleki N, Becerra L, McEwen B. Understanding migraine through the lens of maladaptive stress responses: A model disease of allostatic load. *Neuron*. 2012;73:219-234.
- 177. Kramer DR, Fujii T, Ohiorhenuan I, Liu CY. Cortical spreading depolarization: Pathophysiology, implications, and future directions. *J Clin Neurosci.* 2016;24:22-27.
- 178. Bigal ME, Serrano D, Buse D, et al. Acute migraine medications and evolution from episodic to chronic migraine: A longitudinal population-based study. *Headache*. 2008;48:1157-1168.
- 179. Ba'albaki H, Rapoport A. Mast cells activate the renin angiotensin system and contribute to migraine: A hypothesis. *Headache*. 2008;48:1499-1505.
- 180. Chan SHH, Chan JYH. Angiotensin-generated reactive oxygen species in brain and pathogenesis of cardiovascular diseases. *Antioxid Redox Signal*. 2013;19:1074-1084.
- 181. Zeller JA, Frahm K, Baron R, Stingele R, Deuschl G. Platelet-leukocyte interaction and platelet activation in migraine: A link to ischemic stroke? *J Neurol Neurosurg Psychiatry*. 2004;75:984-987.
- 182. Corrigan F, Mander KA, Leonard AV, Vink R. Neurogenic inflammation after traumatic brain injury and its potentiation of classical inflammation. *J Neuroinflammation*. 2016;13:264.
- 183. Hooper C, Taylor DL, Pocock JM. Pure albumin is a potent trigger of calcium signaling and proliferation in microglia but not macrophages or astrocytes. *J Neurochem.* 2005;92: 1363-1376.
- 184. Ralay Ranaivo H, Wainwright MS. Albumin activates astrocytes and microglia through mitogen-

- activated protein kinase pathways. *Brain Res.* 2010; 1313:222-231.
- 185. Smillie S-J, King R, Kodji X, et al. An ongoing role of α-calcitonin gene-related peptide as part of a protective network against hypertension, vascular hypertrophy, and oxidative stress. *Hypertension*. 2014;63:1056-1062.
- 186. Watts SW. Oh, the places you'll go! My many colored serotonin (apologies to Dr. Suess). *Am J Physiol Heart Circ Physiol.* 2016;311:H1225-H1233.
- 187. Brito R, Castillo G, González J, Valls N, Rodrigo R. Oxidative stress in hypertension: Mechanisms and therapeutic opportunities. *Exp Clin Endocrinol Diabetes*. 2015;123:325-335.
- 188. Chen Q, Wang Q, Zhu J, Xiao Q, Zhang L. Reactive oxygen species: Key regulators in vascular health and diseases. *Br J Pharmacol*. In press; doi: 10.1111/bph.13828.
- 189. Förstermann U, Xia N, Li H. Roles of vascular oxidative stress and nitric oxide in the pathogenesis of atherosclerosis. *Circ Res.* 2017;120:713-735.
- 190. Barbanti P, Aurilia C, Fofi L, Egeo G, Ferroni P. The role of anti-CGRP antibodies in the pathophysiology of primary headaches. *Neurol Sci.* 2017;38: s31-s35.
- 191. MaassenVanDenBrink A, Meijer J, Villalón CM, Ferrari MD. Wiping out CGRP: Potential cardiovascular risks. *Trends Pharmacol Sci.* 2016;37:779-788.
- 192. Schiller F. The migraine tradition. *Bull Hist Med.* 1975;49:1-19.
- 193. Borkum JM. *Chronic Headaches: Biology, Psy-chology, and Behavioral Treatment.* Mahwah, NJ: Lawrence Erlbaum Associates; 2007.